

REVIEW

The regulation of food intake by the gut-brain axis: implications for obesity

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Our understanding of the regulation of appetite has improved considerably over the last few decades. Recent work, stimulated by efforts aimed at curbing the current obesity epidemic, has unravelled some of the complex pathways regulating energy homeostasis. Key factors to this progress have been the discovery of leptin and the neuronal circuitry involved in mediating its effects, as well as the identification of gut hormones that have important physiological roles relating to energy homeostasis. Despite these advances in research, there are currently no effective treatments for the growing problem of obesity. In this article, we summarise the regulatory pathways controlling appetite with a special focus on gut hormones. We detail how recent findings have contributed to our knowledge regarding the pathogenesis and treatment of common obesity. A number of barriers still need to be overcome to develop safe and effective anti-obesity treatments. We outline problems highlighted by historical failures and discuss the potential of augmenting natural satiety signals, such as gut hormones, to treat obesity.

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INTRODUCTION

Over 60 years ago, while studying the effect of hypothalamic lesions on appetite, Brobeck¹ contemplated 'Why does the normal animal eat food? What determines how much it will eat? What changes in the internal environment set the animal to eating? and what changes are associated with satiety?'. The wealth of knowledge gained by recent research has revealed some of the complex control mechanisms relating to Brobeck's questions. Our simplistic views regarding appetite control are constantly being updated to accommodate the complexity of a diverse homeostatic system, which is influenced by a variety of stimuli in the modern world.²

Most of our knowledge regarding appetite control has stemmed from studying the genetics of obesity and altering signalling pathways implicated in obesity to investigate their impact on appetite and body weight regulation. The brain has emerged as the key regulator of appetite. It receives and integrates a variety of homeostatic signals regarding energy status with environmental, social and hedonistic influences to produce an overall response of hunger, which promotes food intake, or satiety, which limits food intake (Figure 1).^{3,4} Although common obesity is unlikely to be a result of pre-existing pathological changes in these circuits, it may be propagated by inappropriate homeostatic responses to energy excesses in an obesogenic environment. Weight gain does not necessarily lead to diminished energy intake and weight loss through dieting is difficult to achieve and sustain. Therefore, this complex regulatory system controlling appetite and body weight is more responsive to a loss in energy stores and is tolerant towards energy gains. Work over the recent years has highlighted a number of redundancies in the appetite regulation system.⁵

These redundancies are likely to contribute to the difficulty in achieving and sustaining meaningful weight loss on anti-obesity therapies.⁶

With energy availability being integral to our survival, it is not surprising that we have powerful hunger signals that are aimed at constantly replenishing and building energy reserves. However, the role for signals that limit food intake is less obvious. It is likely that short-term satiety signals, activated following the intake of a meal and emanating largely from the gut, improve digestive efficiency, nutrient utilisation and prevent large fluctuations in circulating nutrients by preventing consumption of large meals through their effects on the brain. What is evident is that these gut satiety signals are weak, especially in the current obesogenic environment where they are not powerful enough to limit excess energy intake. Nevertheless, understanding these signals and augmenting them pharmacologically may provide an important avenue to treat obesity.

At present, we are unable to tackle the growing obesity problem effectively. Currently, only one drug is licensed for the long-term treatment of obesity.⁷ Despite its efficacy, the costs of and risks from bariatric surgery make it an impractical solution for this worldwide problem. In this review, we summarise our current understanding regarding the central nervous system (CNS) pathways regulating appetite and focus on the role of gut signals in this regard. We discuss the problems with current approaches to tackle obesity, lessons learnt from previous experiences and how our current knowledge may improve the future treatment of obesity. Given the breadth and size of this topic, it is difficult to present a comprehensive account of all the control mechanisms involved. Our eclectic efforts should not diminish the importance of processes that are not discussed in this review.

THE HYPOTHALAMIC CONTROL OF APPETITE

The hypothalamus has a prominent role in coordinating the central control of appetite.⁶ Neural, nutrient and hormonal signals converge directly and indirectly on the hypothalamus forming a network of communication between the gut, pancreas, liver, adipose tissue, brainstem and hypothalamus (Figure 1). The hypothalamus integrates these peripheral signals and modulates appetite in response to them via higher cortical centres, pertaining to food memory and reward, sympathetic and parasympathetic nervous system, influencing gastric motility and hormone secretion among other processes relevant to energy homeostasis.

Within the hypothalamus, the arcuate nucleus (ARC) is one of the main nuclei regulating appetite.⁸ A semipermeable blood-brain barrier resulting from a highly fenestrated local capillary network is thought to allow peripheral signals, such as hormones and nutrients, to gain access to the CNS.^{9,10} The ARC neuronal populations communicate with other hypothalamic nuclei implicated in the control of food intake, such as paraventricular nucleus, dorsomedial nucleus, lateral hypothalamus and ventromedial nucleus.^{11,12} The involvement of specific hypothalamic neuronal populations, especially in the ARC, in appetite regulation has been highlighted in a number of studies. Two well-characterised neuronal populations within the ARC are known to regulate food intake. These are the orexigenic neuropeptide Y/agouti-related peptide neurons and anorexigenic pro-opiomelanocortin/cocaine- and amphetamine-regulated transcript containing neurons.¹³ Neuronal populations in other hypothalamic nuclei involved in coordinating food intake include orexigenic orexin and melanin concentrating hormone releasing neurons of the lateral hypothalamus, anorexigenic brain-derived neurotrophic factor and steroidogenic factor-1 releasing neurons in the ventromedial nucleus.^{14–18}

The discovery of leptin's role in energy homeostasis along with the use of specific genetic manipulations has provided further insight into the specific role of neuronal populations, receptors and cellular pathways involved in altering energy homeostasis. It is now well regarded that these appetite modulating hypothalamic neurons functioned as metabolic sensing units that are capable of 'sensing' nutrients and hormones, including gut hormones.^{19–21} Although, the contribution of pre-existing defects in these neuronal pathways to the development and propagation of common obesity is unclear; these studies have helped our understanding of appetite regulatory pathways.

THE BRAINSTEM SENSES ENERGY BALANCE AND MODULATES FEEDING ACTIVITY

Recent evidence has demonstrated that systems outside the hypothalamus are also important in regulating food intake. The caudal brainstem has a crucial role in ingestive behaviour and is regarded as being the second homeostatic integrator controlling food intake.²² The DVC (dorsal vagal complex) in the caudal brainstem facilitates communication between the periphery and hypothalamus to control food intake. Neural, nutrient and hormonal signals from the gastrointestinal tract are sensed in the brainstem by mechanisms analogous to those seen in the hypothalamus.^{23–25} Receptors and signalling systems influenced by gut hormones and implicated in energy homeostasis have been demonstrated in the vagus nerve and brainstem.²⁶ These appetite signals in the brainstem are relayed to the hypothalamus (Figure 1).^{27–29} The hypothalamus integrates these and other signals to generate an efferent signal, which is transmitted via the brainstem to modulate appetite and gastrointestinal function.^{23,30,31} The vagus has an important role in the transmission of afferent and efferent neural signals between gastrointestinal system and nucleus of the tractus solitarius in the DVC. Cessation of these signals results in altered meal patterns.²⁷

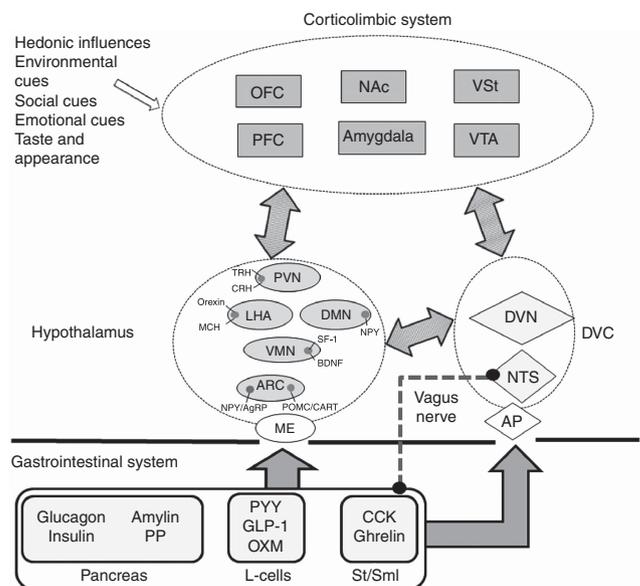


Figure 1. A gut-brain axis controls food intake and appetite in response to homeostatic and non-homeostatic signals. The hypothalamus, brainstem and corticolimbic system form a complex neuronal network, which allows signals from the gut and external environment to be integrated. In response to chemical, mechanical, hormonal and nutritive changes, neural signals are conveyed from the gut to the brainstem via the vagus nerve. Gut hormone signals are also relayed directly to the hypothalamus and brainstem via the median eminence and area postrema, respectively, which forms an incomplete blood-brain barrier. Hedonistic, environmental, social and taste influences are processed via the corticolimbic system and modulate hypothalamic appetite centres. Integration of these signals by the brain produces overall feelings of hunger and satiety. AP, area postrema; ARC, arcuate nucleus; BDNF, brain-derived neurotrophic factor; CCK, cholecystokinin; CRH, corticotropin-releasing hormone; DMN, dorsomedial nucleus; DVC, dorsoventral vagal complex; DVN, dorsoventral neurons; GLP-1, glucagon like peptide-1; L-cells, enteroendocrine L cells of the gastrointestinal tract; LHA, lateral hypothalamic area; MCH, melanin-concentrating hormone; ME, median eminence; NAC, nucleus accumbens; NTS, nucleus of the tractus solitarius; NPY/AgRP, neuropeptide Y and agouti-related peptide; OFC, orbitofrontal cortex; OXM, oxyntomodulin; PFC, pre-frontal cortex; POMC/CART, pro-opiomelanocortin and cocaine- and amphetamine-regulated transcript; PVN, paraventricular nucleus; PP, pancreatic polypeptide; PYY, peptide YY; SF-1, steroidogenic factor-1; St/Sml, stomach and small intestine; TRH, thyrotropin-releasing hormone; VMN, ventromedial nucleus; VSt, ventral striatum; VTA, ventral tegmental area.

THE CORTICOLIMBIC SYSTEM ALLOWS LEARNING, LIKING AND WANTING TO BE INTEGRATED WITH HOMEOSTATIC MECHANISMS

The homeostatic control of food intake is strongly influenced by hedonism, reward and mnemonic representations of food experiences.⁴ These non-homeostatic factors are affected by the environment and processed via the corticolimbic system.

The corticolimbic system structures consist of structures including prefrontal cortex, nucleus accumbens, ventral striatum, hippocampus and amygdala. Histochemical and pharmacological studies, in the corticolimbic and hypothalamic systems, highlight the importance of connections between homeostatic and non-homeostatic centres in altering food intake.⁴ These studies suggest that homeostatic appetite control systems in the hypothalamus are modulated by non-homeostatic influences from the corticolimbic system, allowing environmental cues to dominate homeostatic regulation.⁴ Recent data propose that non-homeostatic systems may also be influenced by homeostatic signals, such as gut hormones.^{32,33} This opens the possibility of

influencing both homeostatic and non-homeostatic systems in an obesogenic environment using single agents such as gut hormones. It has also been hypothesised that altered sensing of these homeostatic signals in common obesity may lead to altered motivation to consume foods.³⁴ Further work is needed to explore the cross-talk between the homeostatic and non-homeostatic systems.

ADIPOSIY SIGNALS MODULATE APPETITE BUT ARE WEAKENED IN OBESITY

The role of leptin and, to some extent, insulin as adiposity feedback signals influencing appetite has been demonstrated and is reviewed in detail elsewhere.³⁵ Resistance to the effects of adiposity signals, such as leptin and insulin, is an important feature of common obesity.³⁵ Leptin levels are elevated in obesity, with impaired hypothalamic leptin signalling, and fall on weight loss. It is unclear if resistance to leptin signalling and altered association of leptin with food intake and satiety are a cause or consequence of obesity. In post-obese subjects, increased post-prandial mean leptin levels are associated with increased post-meal satiety scores as compared with controls, suggesting that the development of obesity may lead to a change in leptin sensitivity.³⁶ Regardless of its aetiology, resistance to adiposity signals may contribute towards increased food intake despite excessive adiposity in obesity, difficulty in achieving weight loss and robust hunger responses elicited after weight loss in obese individuals.

SIGNALS FROM THE GUT INFORM THE BRAIN REGARDING ENERGY STATES

Food intake is the main source of energy in animals. Therefore, it is not surprising that one of the main organs responsible for intake and absorption of food has a key role in informing the brain regarding energy status and altering appetite. The gut-brain axis modulates short-term satiety and hunger responses to regulate the delivery of nutrients and transit of nutrients through the gastrointestinal tract, enabling efficient digestion and storage of energy (Figure 1). This axis also has a role in the regulation of blood glucose levels, adipocyte function and energy expenditure, ensuring maintenance of energy homeostasis following a meal.³⁷

NEURAL SIGNALS FROM THE GUT TO THE BRAINSTEM

Gustatory fibres of cranial nerves VII, IX and X, olfactory fibres of cranial nerve I, and sensory fibres of cranial nerve V are directly or indirectly relayed to the DVC and corticolimbic system.³⁸ Afferent vagus nerve mechanoreceptors, sensitive to gastric and gut intraluminal distension, and chemoreceptors, sensitive to a variety of intraluminal chemical stimuli, relay information from the gastrointestinal tract to the DVC regarding the chemical, mechanical and nutritive properties of ingested food.³⁹

GUT HORMONES

A number of gut hormones have been identified in the gastrointestinal system. They have a fundamental role in coordinating digestive process within the gastrointestinal system via autocrine and paracrine effects but also exert endocrine effects on other organ systems particularly the brain, where some of them have also been found to exist as neurotransmitters. Their role in food intake is reviewed here.

Cholecystokinin

Cholecystokinin was the first gut hormone shown to have a role in appetite regulation.^{40,41} It is secreted post-prandially from I cells of the small intestine and promotes fat and protein digestion.⁴²⁻⁴⁴ Its effects on increasing satiety following meal ingestion are via the CCK1 (cholecystokinin 1) receptors on the vagal afferents,

brainstem and the hypothalamus.⁴⁵⁻⁴⁷ Support for this comes from the hyperphagic and obese Otsuka Long-Evans Tokushima Fatty rat, which lacks the CCK1 receptor.⁴⁸ In contrast, CCK1 knockout mice do not display the same phenotype and the reasons for this are not entirely clear.⁴⁹

Despite the convincing support for cholecystokinin's role in mediating post-prandial satiety, its potential as a therapeutic agent for obesity is limited. Repeated or chronic administration of cholecystokinin is ineffective in reducing long-term food intake.⁵⁰⁻⁵² It is possible that tachyphylaxis or short duration of cholecystokinin's anorectic effects may account for its failure in mediating long-term changes in food intake and weight loss.

Ghrelin

The 'hunger hormone' ghrelin is the only known orexigenic gut hormone.⁵³ It was first identified as an endogenous ligand to the growth hormone secretagogue receptor in the stomach.⁵⁴ It is secreted by the A cells of the gastric fundus and results in increased gastric motility, decreased fat utilisation and stimulation of growth hormone release.^{53,55}

Plasma ghrelin levels are elevated before meals and decline post-prandially in proportion to calories ingested, supporting a role relevant to food intake.⁵⁶⁻⁵⁸ Peripheral administration of ghrelin increases food intake in both rodents and humans.^{59,60} These findings have been replicated in several studies confirming ghrelin as a potent appetite stimulator that has a role in meal initiation. Ghrelin mediates its effects on appetite via the growth hormone secretagogue receptor on vagal afferents and in the ARC to alter neuronal activity of neuropeptide Y/agouti-related peptide neurons.⁶¹ Ghrelin's effects occur in other parts of the hypothalamus and brainstem as well.⁶² A recent report suggested that it has a role in stress induced food reward.⁶³

Ghrelin levels are inversely correlated with bodyweight and rise after weight loss.^{64,65} Furthermore, ghrelin resistance in rodents on high-fat diets has also been described.⁶⁶ These observations suggest that ghrelin may not be important in initiating feeding in obesity.⁵⁵ However, the rise in ghrelin following weight loss may contribute to the poor dietary adherence and regain of lost weight in weight-loss interventions. Strategies to antagonise ghrelin either directly or by inhibition of ghrelin O-acyltransferase (an enzyme required to permit ghrelin to bind to growth hormone secretagogue receptor) are being developed as anti-obesity agents. This approach may lead to more fruitful weight-loss outcomes; however, a successful therapy is yet to emerge.

Peptide YY

Peptide YY (PYY) is a member of the pancreatic polypeptide (PP)-fold family, which also includes neuropeptide Y and PP. These related peptides have a similar PP-fold structural motif and bind to Y family of receptors. PYY is secreted by the L cells of the gastrointestinal tract. PYY₃₋₃₆ is the active form of PYY and binds preferentially to the Y₂ receptor. Its levels are low in the fasting state and rise post-prandially, for several hours, in proportion to calories ingested and especially after a protein-rich meal.⁶⁷⁻⁶⁹ Blunted post-prandial rises in PYY are noted in obesity.⁶⁸ Exaggerated post-prandial PYY rises occur following Roux-en-Y gastric bypass (RYGB).⁷⁰ This increased response may have an important role in the initial weight loss noted by this effective surgical treatment for obesity.⁷¹ The anorectic potential of peripherally administered PYY₃₋₃₆ has been demonstrated and confirmed in several rodent and human studies.^{26,72-74} These anorectic effects of PYY₃₋₃₆ are mediated via the hypothalamus through direct effects on Y₂ receptors in the ARC and indirect effects via the Y₂ receptors on the vagus.²⁶ The net effect of this is to alter ARC neuronal activity.⁷⁵ Decreased gastric emptying by PYY may also have a part in promoting satiety. PYY is one of the hormones considered to mediate the 'ileal brake reflex'. This reflex

results in inhibition of proximal intestine and gastric motor activity with nutrient stimulation of the ileum. This reflex may serve to protect the distal intestine from large nutrient loads. The anorectic potential of PYY₃₋₃₆ has stimulated considerable interest in developing therapies targeting Y₂ receptors for the treatment of obesity.

Pancreatic polypeptide

PP belongs to the PP-fold family of peptides. It is released post-prandially by PP cells in the pancreatic islets and binds to the Y₄ receptor family.²⁶ Its highest affinity is to the Y₄ receptor subtype.⁷⁶ PP delays gastric emptying and also reduces appetite in rodents and humans.⁷⁷⁻⁷⁹ It has been suggested that PP exerts its effects via Y₄ receptors in the ARC, area postrema in the brainstem and vagus.²⁶ Evidence for its central mode of action via hypothalamic centres implicated in energy homeostasis is further strengthened by functional imaging using manganese-enhanced magnetic resonance imaging following PP administration in mice.⁸⁰

Glucagon-like peptide-1

Glucagon-like peptide-1 (GLP-1) is formed from cleavage of the proglucagon precursor.⁸¹ It is secreted by the L cells of the gastrointestinal tract post-prandially in proportion to calories ingested and especially by glucose.⁸² The secretion of GLP-1 from the L cells is stimulated by enteric neuronal signals, as well as via the G protein-coupled 119 receptor, which comes into direct contact with gut luminal contents.^{83,84} GLP-1₇₋₃₆ and GLP-1₇₋₃₇ amide are the major bioactive forms of GLP-1 in the circulation. GLP-1 is degraded by the enzyme dipeptidyl peptidase-4.

GLP-1's incretin effects include increased glucose-dependent insulin release, decreased glucagon secretion and decreased gastric emptying.⁸⁵ Therefore, it has important effects on attenuating post-prandial glucose rise. GLP-1 is also one of the components of the 'ileal brake reflex' and has an anorectic effect on appetite.⁸⁶ Peripheral and central administration in animals and peripheral administration in humans results in reduced food intake.²⁶ GLP-1 mediates this effect via GLP-1 receptors in the ARC, paraventricular nucleus and superior optic nuclei of the hypothalamus, nucleus of the tractus solitarius and area postrema of the brainstem and vagus.^{26,87}

GLP-1's incretin properties have been utilised for the treatment of type 2 diabetes. Exenatide (exendin-4) is a GLP-1 receptor agonist, originally isolated from the saliva of *Heloderma suspectum*, the Gila monster lizard.⁸⁸ It is resistant to dipeptidyl peptidase-4 cleavage. Its use in type 2 diabetes has led to improved glycaemic control as well as weight loss.⁸⁹ Other longer acting dipeptidyl peptidase-4-resistant analogues have been developed and are currently in trials for the treatment of obesity. In a recent study 2-year study, Liraglutide, a once-daily GLP-1 analogue, demonstrated sustained weight loss, increased efficacy compared with orlistat, improvement in metabolic risk factors without any major safety concerns.⁹⁰ Phase III trials for this promising anti-obesity agent are being undertaken.

A recent meta-analysis provides convincing evidence that GLP-1 receptor agonists leads to weight loss in obese patients with or without type 2 diabetes.⁹¹ Furthermore, this meta-analysis also highlighted beneficial effects on blood pressure and total cholesterol. It is unclear if these benefits are a direct result of weight loss or whether there are possible off target effects on GLP-1 receptors present in the vascular system or CNS areas that are not directly involved in appetite control.^{92,93}

Oxyntomodulin

Like GLP-1, oxyntomodulin is a product of proglucagon precursor and is also secreted post-prandially by the L cell in proportion to caloric intake.⁹⁴ Oxyntomodulin exerts its effects mainly via the GLP-1 receptor, although it also has weak affinity to

the glucagon receptor.^{26,95} Oxyntomodulin has a 50 × fold lower affinity for the GLP-1 receptor as compared with GLP-1. Despite its lower receptor affinity, oxyntomodulin reduces food intake and body weight to a similar degree as GLP-1 in rats.²⁶ Oxyntomodulin also delays gastric emptying.⁹⁶ However, unlike GLP-1, oxyntomodulin increases energy expenditure and suppresses ghrelin.^{26,97,98} It is also likely that oxyntomodulin has different pharmacological properties and different tissue-specific actions as compared with GLP-1. The anorectic effects of intraperitoneal oxyntomodulin, but not GLP-1, are inhibited by exendin₉₋₃₉ injected directly into the ARC.⁹⁹ Therefore, oxyntomodulin may exert its anorectic effects in the hypothalamus via different neuronal sub-populations or an unidentified receptor that may also bind to exendin₉₋₃₉ but not GLP-1. This observation is further strengthened by studying the pattern of neuronal activation following oxyntomodulin or GLP-1 administration in mice using manganese-enhanced magnetic resonance imaging. The pattern of hypothalamic neuronal activation is distinct for these two hormones, suggesting that they act differently to mediate their anorectic effects.⁸⁷ The potential of this anorectic gut hormone as an anti-obesity agent is currently being investigated.

Glucagon

Glucagon is a pancreatic hormone produced from the proglucagon precursor molecule in the α cells of the pancreas. Its effects occur via the glucagon receptor, which is mainly expressed in the liver and kidney, although found in a wide range of other tissues as well.¹⁰⁰ Its main effect is to maintain blood glucose levels during fasting and exercise by promoting hepatic glycogenolysis and gluconeogenesis. However, it also has anorectic properties and promotes satiety. Glucagon levels rise post-prandially, especially after a protein diet.¹⁰¹ Glucagon administration in man and rodents results in a reduction of meal size, food intake and body weight.¹⁰¹⁻¹⁰³ Blockade of glucagon increases meal size.¹⁰⁴ Glucagon's satiating effect is most marked when glucagon is infused in to the hepatic portal vein.¹⁰¹ The vagus nerve has been implicated in transducing glucagon signalling to the brainstem to mediate its satiating effects.¹⁰¹ In addition to its satiating effects, glucagon can also increase energy expenditure.¹⁰⁵

Glucagon's anti-obesity potential is severely limited by its detrimental effects on glucose homeostasis. However, co-agonism of glucagon and GLP-1 receptors decreases food intake and increases energy expenditure with no unfavourable effects on glucose homeostasis in rodents.^{106,107} This raises the possibility of utilising dual agonism of glucagon and GLP-1 in the treatment of obesity. Studies utilising this approach are currently underway.

Amylin

Amylin is co-secreted with insulin by pancreatic β cells. In humans, it binds to AMY receptor subtypes, which are complexes of calcitonin receptors with receptor activity-modifying proteins; however, in rodents amylin receptors have not been well characterised.¹⁰⁸ It inhibits gastric secretion, delays gastric emptying improves post-prandial glucose rises and reduces food intake.²⁶ Its satiety effects occur via activation of the serotonin-histamine-dopaminergic system and is independent of the vagus.¹⁰¹ An analogue of amylin is currently being used as adjunctive therapy in diabetes mellitus. Its use is associated with improved glycaemic control and significant weight loss, prompting consideration for its use as an anti-obesity therapy.¹⁰⁹

Other gut hormones

A number of other gut hormones have been implicated in the control of food intake. Neurotensin, first identified as a CNS neurotransmitter, is largely found in the enteroendocrine N cells of the gastrointestinal tract and regulates a number of digestive processes.¹¹⁰ Neurotensin and xenin, a gut hormone with

structural homology to neurotensin, have been shown to have anorexigenic potential in rodents.^{111,112} However, neurotensin's effect on reducing food intake in rodents does not occur chronically, which limits its potential as an anti-obesity therapy.¹¹³

Obestatin is a novel gut hormone that is derived from the preproghrelin precursor and was considered to have opposite effects on food intake to ghrelin.¹¹⁴ Although several studies demonstrated its anorexigenic potential, other studies did not support this.¹¹⁵ Hence, its effects on energy homeostasis require further validation.

Glucagon-like peptide-2 is another product of preproglucagon cleavage in the intestinal L cells.¹¹⁶ Central administration of glucagon-like peptide-2 into rats reduces food intake.¹¹⁷ Peripheral administration in rodents and humans does not alter energy intake.^{118,119} Therefore, it is unlikely to be of much value as an anti-obesity agent.

IMPLICATIONS FOR TREATMENT OF OBESITY: WHY HAVE CURRENT THERAPIES FAILED?

Lifestyle interventions are the cornerstone of obesity management. These interventions include dietary therapy, exercise and behaviour modification. They can achieve a weight loss of up to 10% bodyweight.¹²⁰ However poor adherence and regain of lost weight, due to relapse of lifestyle factors, lead to substantial weight regain. Dropout rates for diets range from 35 to 50% within a year and are predominantly due to the diet becoming hard to follow and frustration at inability to lose further weight.¹²¹ Patients treated through lifestyle measures tend to regain 30 to 35% of their lost weight within a year.¹²⁰ These issues are a familiar problem in obesity management and highlight a homeostatic system that is engineered to defend weight losses with robust hunger responses and diminished energy expenditure. Environmental stimuli and lack of reward on stringent diets make lifestyle interventions even harder to follow. Studies in rodents have highlighted decreased reward value from palatable foods in obesity, leading to increased compulsive eating of palatable foods.¹²² The dopamine system has been implicated in this addiction like adaptation to palatable foods in obesity. Therefore, difficulties in losing further weight, poor perception of satiety after weight loss and diminished reward on dietary therapies are unfortunately to be expected with weight-loss interventions.

At present, orlistat is the only available drug for the long-term treatment of obesity. When combined with lifestyle interventions, it results in a mild supplementary weight loss of 2.9kg above placebo.¹²³ However, no further weight loss is noted thereafter, again reflecting the emergence of multiple compensatory mechanisms, which diminish weight-loss strategies.¹²⁴ Moreover, as for other interventions, there is a marked regain of weight following discontinuation of treatment. Orlistat causes a number of problematic gastrointestinal side effects. These side effects along with limited efficacy and possible cost implications lead to substantial discontinuation of therapy. In trial settings this may be 15–30%, but in non-trial settings up to 98% has been reported.¹²⁵ This further underscores its efficacy as an anti-obesity agent.

DIFFICULTIES IN DRUG DEVELOPMENT IN OBESITY

The above problems highlight the need for drugs that can target the multiple compensatory mechanisms counteracting the effects of weight loss. Although a number of potential targets modulating satiety, hunger, reward and energy expenditure have emerged, developing a safe and effective drug for weight loss remains a big challenge.

Problems with demonstrating safety

Neurotransmitters involved in the neuroendocrine regulation of energy homeostasis are spatially distributed in the CNS and peripheral system. Therefore, it is not surprising that altering

neurotransmitter signalling in a widespread system may lead to unwanted side effects. The use of amphetamine derivatives fenfluramine and dexfenfluramine highlight concerns regarding this. These serotonergic agents lacked the psychostimulant properties associated with amphetamines but were effective in promoting substantial weight loss.¹²⁶ This efficacy, coupled with FDA (Food and Drug Administration, US) approval for short-term and longer term use of these agents, lead to widespread prescribing and off-label use of products containing these agents in the 1990s. The emergence of cardiac valvulopathy resembling serotonin mediated carcinoid syndrome caused a grave concern given its prevalent use.¹²⁷ This prompted investigations into its role in valvulopathies and 20 years after its introduction the FDA estimated a 32.8% incidence of valvular heart disease in those taking fenfluramine. Despite initial warnings issued by the FDA in 1997, these drugs were often not discontinued. By the time it was withdrawn later that year, over 4 million individuals were exposed to fenfluramine and 2 million to dexfenfluramine. Massive litigation eventually culminated in a \$5 billion settlement by Wyeth to 475 000 individuals claiming compensation.

Another example is Rimonabant, a selective CB1 receptor antagonist. This drug emerged on the European market in 2006 after promising initial results and was used extensively as a treatment for obesity.¹²⁸ Post-marketing surveillance resulting from its use in obesity indicated increased psychiatric adverse effects, such as depression and suicide.¹²⁹ After issuing an advisory in 2008, rimonabant was eventually withdrawn from the European market in 2009.

Sibutramine (Reductil), a serotonin and noradrenalin reuptake inhibitor, was licensed for the treatment of obesity in 1997. Cardiovascular concerns emerged because of its sympathomimetic effects and it was noted to increase blood pressure and pulse rate in some patients.¹³⁰ Hence, it was not indicated for patients with a history of cardiovascular disease. Following the emergence of data from Sibutramine Cardiovascular Outcome Trial (SCOUT), it was withdrawn from the European market by the European Medicines Agency.¹³¹ This large study showed a 16% increased rate of serious non-fatal cardiovascular events, such as stroke or myocardial infarction, with sibutramine as compared with placebo. Under pressure from the FDA, this drug was eventually withdrawn from the US market by its manufacturers.

These examples illustrate the potential of adverse events from agents modulating neurotransmission. Subtle and unpredictable side effects from these agents may only become apparent in large, long-term studies. In light of these historical concerns, the FDA has adopted a rigorous approach on approving emerging drugs for the treatment of obesity. In 2010, the FDA rejected Phentermine/Topiramate (Qnexa-Vivus Inc., Mountain View, CA, USA), Bupropion/Naltrexone (Contrave-Orexigen Therapeutics, La Jolla, CA, USA) and Lorcaserin Hydrochloride (Lorcaserin-Arena Pharmaceuticals, San Diego, CA, USA) primarily over safety concerns. Both Lorcaserin and Phentermine/Topiramate have refilled their applications to the FDA, with Phentermine/Topiramate being recommended for approval by an FDA advisory panel earlier this year. Orexigen is attempting to address the cardiovascular safety concerns highlighted by the FDA for Bupropion/Naltrexone by conducting a cardiovascular outcomes trial. Although these potential anti-obesity drugs may eventually obtain approval for their use, such stringency by drug regulatory authorities, on a background of previous failures, could be a potential deterrent for pharmaceutical companies looking to develop new anti-obesity agents.

The complexity of the biological systems regulating energy homeostasis

The discovery of leptin as an adiposity factor involved in appetite regulation in 1994 was heralded by many as a finding which

would revolutionise the treatment of obesity. Amgen Inc. (Thousand Oaks, CA, USA) embraced this discovery, investing millions with the hope of using leptin as an anti-obesity agent. However, most human trials with leptin in common obesity have been disappointing. Over the last decade, and as mentioned earlier, it has become increasingly apparent that obesity is a leptin-resistant state with hyperleptinaemia. Therefore, simply replacing leptin may not be a useful strategy to reduce appetite in obese individuals. The mechanisms regulating energy homeostasis and metabolism are complex. Our current views regarding these processes are often too simplistic to predict the long-term efficacy and consequences of altering biological systems in obesity.

Unexpected biological outcomes

The emergence of ciliary neurotrophic factor as a nerve growth factor involved in producing an anorectic response was also very promising. In trials for the treatment of motor neuron disease, ciliary neurotrophic factor caused marked weight loss. Subsequent investigations highlighted that ciliary neurotrophic factor may mediate its effect on energy homeostasis by stimulating increased neurogenesis in pathways linked to energy homeostasis, resulting in increased leptin responsive neurons and leptin-like signalling.^{132,133} This effect was preserved in leptin-resistant obesity states raising hopes that this may be an effective treatment in common obesity. Axokine (Regeneron Pharmaceuticals Inc., Tarrytown, NY, USA), a modified version of ciliary neurotrophic factor, was investigated as a treatment for weight loss. Initial results were encouraging.¹³⁴ However, a phase III study resulted in the development of neutralising antibodies in 70% of participants after 3 months of therapy, compromising its efficacy and raising concerns about disrupting neuronal regeneration processes in individuals treated with this agent. These scenarios are a common concern when translating successes from the lab to patients. It is difficult to assess the biological responses of potential anti-obesity agents comprehensively in rodents and small duration, short-term studies in humans.

WHERE NEXT?

Lessons from an effective but invasive and costly treatment

Bariatric surgical procedures are currently the most effective weight-loss treatment for obesity. Weight loss, sustainance of weight decline, incidence of obesity-related diseases and quality of life significantly improve with bariatric surgery, as compared with other conventional therapies.¹³⁵ There are several type of bariatric surgical procedures. These include restrictive procedures such as gastric banding, malabsorptive procedures such as biliopancreatic diversion and mixed procedures such as RYGB that restrict caloric intake and incorporate an element of gastrointestinal bypass. Appetite, weight loss and amelioration of type 2 diabetes are more effective with mixed procedures such as RYGB. A rise in post-prandial gut hormone secretion is noted immediately after duodenal switch in RYGB surgery.⁷⁰ This may account for the reduced hunger and altered food preference, despite substantial weight loss, in patients undergoing RYGB surgery. Unfortunately, the cost, operative risks and limited availability of the procedure, even in developed countries, makes bariatric surgery an impractical option for the treatment of obesity. This form of treatment is usually reserved for severely obese, particularly those with obesity exacerbated health conditions. However, they illustrate the therapeutic potential of altering gut signals in managing obesity. Hence, mimicking this model, by using these naturally occurring satiety signals as anti-obesity treatments, is likely to be a successful strategy to combat obesity and may circumvent safety problems associated with past failures in obesity therapy.

Combination therapy

Given the redundancies in the appetite regulation pathways and multiple compensatory mechanisms elicited on weight loss, combination therapy using multiple agents may offer better weight-loss outcomes. Furthermore, this strategy may also allow lower doses for each agent to be used, thereby reducing the emergence of potential side effects. This rationale has been used in the treatment of other common complex diseases, such as hypertension and diabetes. Bupropion/naltrexone (Contrave) and phentermine/ topiramate (Qnexa), discussed earlier in this review, as well as bupropion/zonisamide (Empatic) are based on this approach.¹³⁶ Early results from combinatorial gut hormone approaches using GLP-1 receptor agonists and PYY₃₋₃₆ show additive weight-loss benefits in rodents and short human studies.^{33,137,138} Other combination gut hormone approaches that are currently being investigated include PP/PYY₃₋₃₆ (Obineptide), GLP-1/Glucagon co-agonism and metreleptin/pramlintide.¹³⁶

CONCLUSIONS

Metabolism and continuous availability of energy is integral to our survival. The gut-brain axis represents a holistic control system where the peripheral and central systems link and feedback to each other continuously to control energy homeostasis. This cross-talk allows us to adapt to changes in the environment and maintain energy homeostasis; ensuring energy is constantly available for the body's needs. Our perception of hunger and satiety is modulated through this network depending on energy status, environmental stimuli, as well as other needs and behaviours.

The sensitivity and nature of these signals controlling energy homeostasis is biased towards promoting weight gain and preventing weight loss. Modulating these signals by pharmacological means would help tackle the obesity problem. So-far efforts in this regard have been unsuccessful. The complexity, diversity and redundancy of pathways regulating food intake present a number of obstacles to effective and safe drug development. Nevertheless, our efforts to delineate the mechanisms that control food intake, as well as lessons from failures and successes, have provided an important framework that will enable us to identify and study targets for drug development. Gut hormones have been shown to have a fundamental role in energy homeostasis. The use of gut hormones as anti-obesity treatments is an attractive option and shows considerable promise. Although it is unlikely that we will ever have a miracle cure to a problem that has societal and environmental origins, judicious use of combinatorial gut hormone regimens is likely to help our currently futile efforts to tackle this growing problem.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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REFERENCES

- 1 Brobeck JR. Mechanism of the development of obesity in animals with hypothalamic lesions. *Physiol Rev* 1946; **26**: 541–559.
- 2 Speakman JR, Levitsky DA, Allison DB, Bray MS, de Castro JM, Clegg DJ *et al*. Set points, settling points and some alternative models: theoretical options to

- understand how genes and environments combine to regulate body adiposity. *Dis Model Mech* 2011; **4**: 733–745.
- 3 Schwartz MW, Woods SC, Porte Jr D, Seeley RJ, Baskin DG. Central nervous system control of food intake. *Nature* 2000; **404**: 661–671.
 - 4 Berthoud HR. Homeostatic and non-homeostatic pathways involved in the control of food intake and energy balance. *Obesity (Silver Spring)* 2006; **14**(Suppl 5): 197S–200S.
 - 5 Beck B. KO's and organisation of peptidergic feeding behavior mechanisms. *Neurosci Biobehav Rev* 2001; **25**: 143–158.
 - 6 Schwartz MW. Central nervous system regulation of food intake. *Obesity (Silver Spring)* 2006; **14**(Suppl 1): 1S–8S.
 - 7 Hussain SS, Bloom SR. The pharmacological treatment and management of obesity. *Postgrad Med* 2011; **123**: 34–44.
 - 8 Konner AC, Klockener T, Bruning JC. Control of energy homeostasis by insulin and leptin: targeting the arcuate nucleus and beyond. *Physiol Behav* 2009; **97**: 632–638.
 - 9 Broadwell RD, Brightman MW. Entry of peroxidase into neurons of the central and peripheral nervous systems from extracerebral and cerebral blood. *J Comp Neurol* 1976; **166**: 257–283.
 - 10 Peruzzo B, Pastor FE, Blazquez JL, Schobitz K, Pelaez B, Amat P *et al.* A second look at the barriers of the medial basal hypothalamus. *Exp Brain Res* 2000; **132**: 10–26.
 - 11 Kalra SP, Dube MG, Pu S, Xu B, Horvath TL, Kalra PS. Interacting appetite-regulating pathways in the hypothalamic regulation of body weight. *Endocr Rev* 1999; **20**: 68–100.
 - 12 Bouret SG, Draper SJ, Simerly RB. Formation of projection pathways from the arcuate nucleus of the hypothalamus to hypothalamic regions implicated in the neural control of feeding behavior in mice. *J Neurosci* 2004; **24**: 2797–2805.
 - 13 Cone RD, Cowley MA, Butler AA, Fan W, Marks DL, Low MJ. The arcuate nucleus as a conduit for diverse signals relevant to energy homeostasis. *Int J Obes Relat Metab Disord* 2001; **25**(Suppl 5): S63–S67.
 - 14 Sakurai T, Amemiya A, Ishii M, Matsuzaki I, Chemelli RM, Tanaka H *et al.* Orexins and orexin receptors: a family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. *Cell* 1998; **92**: 573–585.
 - 15 Qu D, Ludwig DS, Gammeltoft S, Piper M, Pelleymounter MA, Cullen MJ *et al.* A role for melanin-concentrating hormone in the central regulation of feeding behaviour. *Nature* 1996; **380**: 243–247.
 - 16 Rossi M, Choi SJ, O'Shea D, Miyoshi T, Ghatei MA, Bloom SR. Melanin-concentrating hormone acutely stimulates feeding, but chronic administration has no effect on body weight. *Endocrinology* 1997; **138**: 351–355.
 - 17 Xu B, Goulding EH, Zang K, Cepoi D, Cone RD, Jones KR *et al.* Brain-derived neurotrophic factor regulates energy balance downstream of melanocortin-4 receptor. *Nat Neurosci* 2003; **6**: 736–742.
 - 18 Dhillon H, Zigman JM, Ye C, Lee CE, McGovern RA, Tang V *et al.* Leptin directly activates SF1 neurons in the VMH, and this action by leptin is required for normal body-weight homeostasis. *Neuron* 2006; **49**: 191–203.
 - 19 Lam TK, Schwartz GJ, Rossetti L. Hypothalamic sensing of fatty acids. *Nat Neurosci* 2005; **8**: 579–584.
 - 20 Jordan SD, Konner AC, Bruning JC. Sensing the fuels: glucose and lipid signaling in the CNS controlling energy homeostasis. *Cell Mol Life Sci* 2010; **67**: 3255–3273.
 - 21 Levin BE, Magnan C, Dunn-Meynell A, Le Foll C. Metabolic sensing and the brain: who, what, where, and how? *Endocrinology* 2011; **152**: 2552–2557.
 - 22 Grill HJ, Kaplan JM. The neuroanatomical axis for control of energy balance. *Front Neuroendocrinol* 2002; **23**: 2–40.
 - 23 Blevins JE, Baskin DG. Hypothalamic-brainstem circuits controlling eating. *Forum Nutr* 2010; **63**: 133–140.
 - 24 Grill HJ, Schwartz MW, Kaplan JM, Foxhall JS, Breininger J, Baskin DG. Evidence that the caudal brainstem is a target for the inhibitory effect of leptin on food intake. *Endocrinology* 2002; **143**: 239–246.
 - 25 Lebrun B, Bariohay B, Moysé E, Jean A. Brain-derived neurotrophic factor (BDNF) and food intake regulation: a minireview. *Auton Neurosci* 2006; **126–127**: 30–38.
 - 26 Chaudhuri O, Small C, Bloom S. Gastrointestinal hormones regulating appetite. *Philos Trans R Soc Lond B Biol Sci* 2006; **361**: 1187–1209.
 - 27 Schwartz GJ. The role of gastrointestinal vagal afferents in the control of food intake: current prospects. *Nutrition* 2000; **16**: 866–873.
 - 28 Price CJ, Hoyda TD, Ferguson AV. The area postrema: a brain monitor and integrator of systemic autonomic state. *Neuroscientist* 2008; **14**: 182–194.
 - 29 Ter Horst GJ, de Boer P, Luiten PG, van Willigen JD. Ascending projections from the solitary tract nucleus to the hypothalamus. A Phaseolus vulgaris lectin tracing study in the rat. *Neuroscience* 1989; **31**: 785–797.
 - 30 ter Horst GJ, Luiten PG, Kuipers F. Descending pathways from hypothalamus to dorsal motor vagus and ambiguus nuclei in the rat. *J Auton Nerv Syst* 1984; **11**: 59–75.
 - 31 Grijalva CV, Novin D. The role of the hypothalamus and dorsal vagal complex in gastrointestinal function and pathophysiology. *Ann NY Acad Sci* 1990; **597**: 207–222.
 - 32 Grill HJ, Skibicka KP, Hayes MR. Imaging obesity: fMRI, food reward, and feeding. *Cell Metab* 2007; **6**: 423–425.
 - 33 De Silva A, Salem V, Long CJ, Makwana A, Newbould RD, Rabiner EA *et al.* The gut hormones PYY(3-36) and GLP-1(7-36 amide) reduce food intake and modulate brain activity in appetite centers in humans. *Cell Metab* 2011; **14**: 700–706.
 - 34 Page KA, Seo D, Belfort-DeAguiar R, Lacadie C, Dzura J, Naik S *et al.* Circulating glucose levels modulate neural control of desire for high-calorie foods in humans. *J Clin Invest* 2011; **121**: 4161–4169.
 - 35 Velloso LA, Schwartz MW. Altered hypothalamic function in diet-induced obesity. *Int J Obes (Lond)* 2011; **35**: 1455–1465.
 - 36 Raben A, Astrup A. Leptin is influenced both by predisposition to obesity and diet composition. *Int J Obes Relat Metab Disord* 2000; **24**: 450–459.
 - 37 Murphy KG, Bloom SR. Gut hormones and the regulation of energy homeostasis. *Nature* 2006; **444**: 854–859.
 - 38 Rolls ET. Brain mechanisms underlying flavour and appetite. *Philos Trans R Soc Lond B Biol Sci* 2006; **361**: 1123–1136.
 - 39 Schwartz GJ. Brainstem integrative function in the central nervous system control of food intake. *Forum Nutr* 2010; **63**: 141–151.
 - 40 Gibbs J, Young RC, Smith GP. Cholecystokinin decreases food intake in rats. *J Comp Physiol Psychol* 1973; **84**: 488–495.
 - 41 Kissileff HR, Pi-Sunyer FX, Thornton J, Smith GP. C-terminal octapeptide of cholecystokinin decreases food intake in man. *Am J Clin Nutr* 1981; **34**: 154–160.
 - 42 Buffa R, Solcia E, Go VL. Immunohistochemical identification of the cholecystokinin cell in the intestinal mucosa. *Gastroenterology* 1976; **70**: 528–532.
 - 43 Liddle RA, Goldfine ID, Rosen MS, Taplitz RA, Williams JA. Cholecystokinin bioactivity in human plasma. Molecular forms, responses to feeding, and relationship to gallbladder contraction. *J Clin Invest* 1985; **75**: 1144–1152.
 - 44 Rehfeld JF, Bungaard JR, Friis-Hansen L, Goetze JP. On the tissue-specific processing of procholecystokinin in the brain and gut—a short review. *J Physiol Pharmacol* 2003; **54**(Suppl 4): 73–79.
 - 45 Moran TH, Ameglio PJ, Schwartz GJ, McHugh PR. Blockade of type A, not type B, CCK receptors attenuates satiety actions of exogenous and endogenous CCK. *Am J Physiol* 1992; **262**(1 Pt 2): R46–R50.
 - 46 Zittel TT, Glatzle J, Kreis ME, Starlinger M, Eichner M, Raybould HE *et al.* C-fos protein expression in the nucleus of the solitary tract correlates with cholecystokinin dose injected and food intake in rats. *Brain Res* 1999; **846**: 1–11.
 - 47 Blevins JE, Stanley BG, Reidelberger RD. Brain regions where cholecystokinin suppresses feeding in rats. *Brain Res* 2000; **860**: 1–10.
 - 48 Moran TH, Katz LF, Plata-Salamán CR, Schwartz GJ. Disordered food intake and obesity in rats lacking cholecystokinin A receptors. *Am J Physiol* 1998; **274**(3 Pt 2): R618–R625.
 - 49 Kopin AS, Mathes WF, McBride EW, Nguyen M, Al-Haider W, Schmitz F *et al.* The cholecystokinin-A receptor mediates inhibition of food intake yet is not essential for the maintenance of body weight. *J Clin Invest* 1999; **103**: 383–391.
 - 50 Crawley JN, Beinfeld MC. Rapid development of tolerance to the behavioural actions of cholecystokinin. *Nature* 1983; **302**: 703–706.
 - 51 West DB, Fey D, Woods SC. Cholecystokinin persistently suppresses meal size but not food intake in free-feeding rats. *Am J Physiol* 1984; **246**(5 Pt 2): R776–R787.
 - 52 West DB, Greenwood MR, Sullivan AC, Prescod L, Marzullo LR, Triscari J. Infusion of cholecystokinin between meals into free-feeding rats fails to prolong the intermeal interval. *Physiol Behav* 1987; **39**: 111–115.
 - 53 Kojima M, Kangawa K. Ghrelin: structure and function. *Physiol Rev* 2005; **85**: 495–522.
 - 54 Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature* 1999; **402**: 656–660.
 - 55 Patterson M, Bloom SR, Gardiner JV. Ghrelin and appetite control in humans—Potential application in the treatment of obesity. *Peptides* 2011; **32**: 2290–2294.
 - 56 Cummings DE, Purnell JQ, Frayo RS, Schmidova K, Wisse BE, Weigle DS. A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. *Diabetes* 2001; **50**: 1714–1719.
 - 57 Tschöp M, Wawarta R, Riepl RL, Friedrich S, Bidlingmaier M, Landgraf R *et al.* Post-prandial decrease of circulating human ghrelin levels. *J Endocrinol Invest* 2001; **24**: RC19–RC21.
 - 58 Ariyasu H, Takaya K, Tagami T, Ogawa Y, Hosoda K, Akamizu T *et al.* Stomach is a major source of circulating ghrelin, and feeding state determines plasma ghrelin-like immunoreactivity levels in humans. *J Clin Endocrinol Metab* 2001; **86**: 4753–4758.
 - 59 Wren AM, Seal LJ, Cohen MA, Brynes AE, Frost GS, Murphy KG *et al.* Ghrelin enhances appetite and increases food intake in humans. *J Clin Endocrinol Metab* 2001; **86**: 5992.
 - 60 Wren AM, Small CJ, Abbott CR, Dhillo WS, Seal LJ, Cohen MA *et al.* Ghrelin causes hyperphagia and obesity in rats. *Diabetes* 2001; **50**: 2540–2547.
 - 61 Castaneda TR, Tong J, Datta R, Culler M, Tschöp MH. Ghrelin in the regulation of body weight and metabolism. *Front Neuroendocrinol* 2010; **31**: 44–60.

- 62 Lawrence CB, Snape AC, Baudoin FM, Luckman SM. Acute central ghrelin and GH secretagogues induce feeding and activate brain appetite centers. *Endocrinology* 2002; **143**: 155–162.
- 63 Chuang JC, Perello M, Sakata I, Osborne-Lawrence S, Savitt JM, Lutter M *et al*. Ghrelin mediates stress-induced food-reward behavior in mice. *J Clin Invest* 2011; **121**: 2684–2692.
- 64 Tschop M, Weyer C, Tataranni PA, Devanarayan V, Ravussin E, Heiman ML. Circulating ghrelin levels are decreased in human obesity. *Diabetes* 2001; **50**: 707–709.
- 65 Cummings DE, Weigle DS, Frayo RS, Breen PA, Ma MK, Dellinger EP *et al*. Plasma ghrelin levels after diet-induced weight loss or gastric bypass surgery. *N Engl J Med* 2002; **346**: 1623–1630.
- 66 Gardiner JV, Campbell D, Patterson M, Kent A, Ghatei MA, Bloom SR *et al*. The hyperphagic effect of ghrelin is inhibited in mice by a diet high in fat. *Gastroenterology* 2010; **138**: 2468–2476, 2476.e1.
- 67 Adrian TE, Ferri GL, Bacarese-Hamilton AJ, Fuessl HS, Polak JM, Bloom SR. Human distribution and release of a putative new gut hormone, peptide YY. *Gastroenterology* 1985; **89**: 1070–1077.
- 68 Batterham RL, Cohen MA, Ellis SM, Le Roux CW, Withers DJ, Frost GS *et al*. Inhibition of food intake in obese subjects by peptide YY3-36. *N Engl J Med* 2003; **349**: 941–948.
- 69 Batterham RL, Heffron H, Kapoor S, Chivers JE, Chandarana K, Herzog H *et al*. Critical role for peptide YY in protein-mediated satiation and body-weight regulation. *Cell Metab* 2006; **4**: 223–233.
- 70 Vincent RP, le Roux CW. Changes in gut hormones after bariatric surgery. *Clin Endocrinol (Oxf)* 2008; **69**: 173–179.
- 71 Chandarana K, Gelegen C, Karra E, Choudhury AI, Drew ME, Fauveau V *et al*. Diet and gastrointestinal bypass-induced weight loss: the roles of ghrelin and peptide YY. *Diabetes* 2011; **60**: 810–818.
- 72 Vrang N, Madsen AN, Tang-Christensen M, Hansen G, Larsen PJ. PYY(3-36) reduces food intake and body weight and improves insulin sensitivity in rodent models of diet-induced obesity. *Am J Physiol Regul Integr Comp Physiol* 2006; **291**: R367–R375.
- 73 Degen L, Oesch S, Casanova M, Graf S, Ketterer S, Drewe J *et al*. Effect of peptide YY3-36 on food intake in humans. *Gastroenterology* 2005; **129**: 1430–1436.
- 74 Sloth B, Holst JJ, Flint A, Gregersen NT, Astrup A. Effects of PYY1-36 and PYY3-36 on appetite, energy intake, energy expenditure, glucose and fat metabolism in obese and lean subjects. *Am J Physiol Endocrinol Metab* 2007; **292**: E1062–E1068.
- 75 Batterham RL, Cowley MA, Small CJ, Herzog H, Cohen MA, Dakin CL *et al*. Gut hormone PYY(3-36) physiologically inhibits food intake. *Nature* 2002; **418**: 650–654.
- 76 Michel MC, Beck-Sickinger A, Cox H, Doods HN, Herzog H, Larhammar D *et al*. XVI. International Union of Pharmacology recommendations for the nomenclature of neuropeptide Y, peptide YY, and pancreatic polypeptide receptors. *Pharmacol Rev* 1998; **50**: 143–150.
- 77 Schmidt PT, Naslund E, Gryback P, Jacobsson H, Holst JJ, Hilsted L *et al*. A role for pancreatic polypeptide in the regulation of gastric emptying and short-term metabolic control. *J Clin Endocrinol Metab* 2005; **90**: 5241–5246.
- 78 Asakawa A, Inui A, Ueno N, Fujimiya M, Fujino MA, Kasuga M. Mouse pancreatic polypeptide modulates food intake, while not influencing anxiety in mice. *Peptides* 1999; **20**: 1445–1448.
- 79 Batterham RL, Le Roux CW, Cohen MA, Park AJ, Ellis SM, Patterson M *et al*. Pancreatic polypeptide reduces appetite and food intake in humans. *J Clin Endocrinol Metab* 2003; **88**: 3989–3992.
- 80 Hankir MK, Parkinson JR, Minnion JS, Addison ML, Bloom SR, Bell JD. Peptide YY 3-36 and pancreatic polypeptide differentially regulate hypothalamic neuronal activity in mice *in vivo* as measured by manganese-enhanced magnetic resonance imaging. *J Neuroendocrinol* 2011; **23**: 371–380.
- 81 Dhanvantari S, Seidah NG, Brubaker PL. Role of prohormone convertases in the tissue-specific processing of proglucagon. *Mol Endocrinol* 1996; **10**: 342–355.
- 82 Herrmann C, Goke R, Richter G, Fehmann HC, Arnold R, Goke B. Glucagon-like peptide-1 and glucose-dependent insulin-releasing polypeptide plasma levels in response to nutrients. *Digestion* 1995; **56**: 117–126.
- 83 Rocca AS, Brubaker PL. Role of the vagus nerve in mediating proximal nutrient-induced glucagon-like peptide-1 secretion. *Endocrinology* 1999; **140**: 1687–1694.
- 84 Chu ZL, Carroll C, Alfonso J, Gutierrez V, He H, Lucman A *et al*. A role for intestinal endocrine cell-expressed G protein-coupled receptor 119 in glycemic control by enhancing glucagon-like peptide-1 and glucose-dependent insulinotropic peptide release. *Endocrinology* 2008; **149**: 2038–2047.
- 85 Kreymann B, Williams G, Ghatei MA, Bloom SR. Glucagon-like peptide-1 7-36: a physiological incretin in man. *Lancet* 1987; **2**: 1300–1304.
- 86 Drucker DJ. The biology of incretin hormones. *Cell Metab* 2006; **3**: 153–165.
- 87 Parkinson JR, Chaudhri OB, Kuo YT, Field BC, Herlihy AH, Dhillon WS *et al*. Differential patterns of neuronal activation in the brainstem and hypothalamus following peripheral injection of GLP-1, oxyntomodulin and lithium chloride in mice detected by manganese-enhanced magnetic resonance imaging (MEMRI). *Neuroimage* 2009; **44**: 1022–1031.
- 88 Eng J, Kleinman WA, Singh L, Raufman JP. Isolation and characterization of exendin-4, an exendin-3 analogue, from *Heloderma suspectum* venom. Further evidence for an exendin receptor on dispersed acini from guinea pig pancreas. *J Biol Chem* 1992; **267**: 7402–7405.
- 89 Shyangdan DS, Royle P, Clar C, Sharma P, Waugh N, Snaith A. Glucagon-like peptide analogues for type 2 diabetes mellitus. *Cochrane Database Syst Rev* 2011; CD006423.
- 90 Astrup A, Carraro R, Finer N, Harper A, Kunesova M, Lean ME *et al*. Safety, tolerability and sustained weight loss over 2 years with the once-daily human GLP-1 analog, liraglutide. *Int J Obes (Lond)* 2011; e-pub ahead of print 16 August 2011; doi:10.1038/ijo.2011.158.
- 91 Vilsboll T, Christensen M, Junker AE, Knop FK, Gluud LL. Effects of glucagon-like peptide-1 receptor agonists on weight loss: systematic review and meta-analyses of randomised controlled trials. *BMJ* 2012; **344**: d7771.
- 92 Arakawa M, Mita T, Azuma K, Ebato C, Goto H, Nomiya T *et al*. Inhibition of monocyte adhesion to endothelial cells and attenuation of atherosclerotic lesion by a glucagon-like peptide-1 receptor agonist, exendin-4. *Diabetes* 2010; **59**: 1030–1037.
- 93 Jin SL, Han VK, Simmons JG, Towle AC, Lauder JM, Lund PK. Distribution of glucagonlike peptide I (GLP-I), glucagon, and glicentin in the rat brain: an immunocytochemical study. *J Comp Neurol* 1988; **271**: 519–532.
- 94 Le Quellec A, Kervran A, Blache P, Ciurana AJ, Bataille D. Oxyntomodulin-like immunoreactivity: diurnal profile of a new potential enterogastrone. *J Clin Endocrinol Metab* 1992; **74**: 1405–1409.
- 95 Baggio LL, Huang Q, Brown TJ, Drucker DJ. Oxyntomodulin and glucagon-like peptide-1 differentially regulate murine food intake and energy expenditure. *Gastroenterology* 2004; **127**: 546–558.
- 96 Schjoldager B, Mortensen PE, Myhre J, Christiansen J, Holst JJ. Oxyntomodulin from distal gut. Role in regulation of gastric and pancreatic functions. *Dig Dis Sci* 1989; **34**: 1411–1419.
- 97 Wynne K, Park AJ, Small CJ, Meeran K, Ghatei MA, Frost GS *et al*. Oxyntomodulin increases energy expenditure in addition to decreasing energy intake in overweight and obese humans: a randomised controlled trial. *Int J Obes (Lond)* 2006; **30**: 1729–1736.
- 98 Chaudhri OB, Parkinson JR, Kuo YT, Druce MR, Herlihy AH, Bell JD *et al*. Differential hypothalamic neuronal activation following peripheral injection of GLP-1 and oxyntomodulin in mice detected by manganese-enhanced magnetic resonance imaging. *Biochem Biophys Res Commun* 2006; **350**: 298–306.
- 99 Dakin CL, Small CJ, Batterham RL, Neary NM, Cohen MA, Patterson M *et al*. Peripheral oxyntomodulin reduces food intake and body weight gain in rats. *Endocrinology* 2004; **145**: 2687–2695.
- 100 Svoboda M, Tastenoy M, Vertongen P, Robberecht P. Relative quantitative analysis of glucagon receptor mRNA in rat tissues. *Mol Cell Endocrinol* 1994; **105**: 131–137.
- 101 Woods SC, Lutz TA, Geary N, Langhans W. Pancreatic signals controlling food intake; insulin, glucagon and amylin. *Philos Trans R Soc Lond B Biol Sci* 2006; **361**: 1219–1235.
- 102 Schulman JL, Carleton JL, Whitney G, Whitehorn JC. Effect of glucagon on food intake and body weight in man. *J Appl Physiol* 1957; **11**: 419–421.
- 103 de Castro JM, Paullin SK, DeLugas GM. Insulin and glucagon as determinants of body weight set point and microregulation in rats. *J Comp Physiol Psychol* 1978; **92**: 571–579.
- 104 Langhans W, Zeiger U, Scharrer E, Geary N. Stimulation of feeding in rats by intraperitoneal injection of antibodies to glucagon. *Science* 1982; **218**: 894–896.
- 105 Habegger KM, Heppner KM, Geary N, Bartness TJ, DiMarchi R, Tschop MH. The metabolic actions of glucagon revisited. *Nat Rev Endocrinol* 2010; **6**: 689–697.
- 106 Poci A, Carrington PE, Adams JR, Wright M, Eiermann G, Zhu L *et al*. Glucagon-like peptide 1/glucagon receptor dual agonism reverses obesity in mice. *Diabetes* 2009; **58**: 2258–2266.
- 107 Day JW, Ottaway N, Patterson JT, Gelfanov V, Smiley D, Gidda J *et al*. A new glucagon and GLP-1 co-agonist eliminates obesity in rodents. *Nat Chem Biol* 2009; **5**: 749–757.
- 108 Bailey RJ, Walker CS, Ferner AH, Loomes KM, Prijic G, Halim A *et al*. Pharmacological characterisation of rat amylin receptors: implications for the identification of amylin receptor subtypes. *Br J Pharmacol* 2011; **Oct**: 20.
- 109 Hollander P, Maggs DG, Ruggles JA, Fineman M, Shen L, Kolterman OG *et al*. Effect of pramlintide on weight in overweight and obese insulin-treated type 2 diabetes patients. *Obes Res* 2004; **12**: 661–668.
- 110 Mustain WC, Rychahou PG, Evers BM. The role of neurotensin in physiologic and pathologic processes. *Curr Opin Endocrinol Diabetes Obes* 2011; **18**: 75–82.
- 111 Cui H, Cai F, Belsham DD. Anorexigenic hormones leptin, insulin, and alpha-melanocyte-stimulating hormone directly induce neurotensin (NT) gene expression in novel NT-expressing cell models. *J Neurosci* 2005; **25**: 9497–9506.

- 112 Kim ER, Leckstrom A, Mizuno TM. Impaired anorectic effect of leptin in neurotensin receptor 1-deficient mice. *Behav Brain Res* 2008; **194**: 66–71.
- 113 Cooke JH, Patterson M, Patel SR, Smith KL, Ghatei MA, Bloom SR *et al*. Peripheral and central administration of xenin and neurotensin suppress food intake in rodents. *Obesity (Silver Spring)* 2009; **17**: 1135–1143.
- 114 Zhang JV, Ren PG, Avsian-Kretchmer O, Luo CW, Rauch R, Klein C *et al*. Obestatin, a peptide encoded by the ghrelin gene, opposes ghrelin's effects on food intake. *Science* 2005; **310**: 996–999.
- 115 Li JB, Asakawa A, Cheng K, Li Y, Chaolu H, Tsai M *et al*. Biological effects of obestatin. *Endocrine* 2011; **39**: 205–211.
- 116 Damholt AB, Buchan AM, Holst JJ, Kofod H. Proglucagon processing profile in canine L cells expressing endogenous prohormone convertase 1/3 and prohormone convertase 2. *Endocrinology* 1999; **140**: 4800–4808.
- 117 Tang-Christensen M, Larsen PJ, Thulesen J, Romer J, Vrang N. The proglucagon-derived peptide, glucagon-like peptide-2, is a neurotransmitter involved in the regulation of food intake. *Nat Med* 2000; **6**: 802–807.
- 118 Scott RB, Kirk D, MacNaughton WK, Meddings JB. GLP-2 augments the adaptive response to massive intestinal resection in rat. *Am J Physiol* 1998; **275**(5 Pt 1): G911–G921.
- 119 Schmidt PT, Naslund E, Gryback P, Jacobsson H, Hartmann B, Holst JJ *et al*. Peripheral administration of GLP-2 to humans has no effect on gastric emptying or satiety. *Regul Pept* 2003; **116**: 21–25.
- 120 Wadden TA, Butryn ML, Wilson C. Lifestyle modification for the management of obesity. *Gastroenterology* 2007; **132**: 2226–2238.
- 121 Alhassan S, Kim S, Bersamin A, King AC, Gardner CD. Dietary adherence and weight loss success among overweight women: results from the A TO Z weight loss study. *Int J Obes (Lond)* 2008; **32**: 985–991.
- 122 Johnson PM, Kenny PJ. Dopamine D2 receptors in addiction-like reward dysfunction and compulsive eating in obese rats. *Nat Neurosci* 2010; **13**: 635–641.
- 123 Rucker D, Padwal R, Li SK, Curioni C, Lau DC. Long term pharmacotherapy for obesity and overweight: updated meta-analysis. *BMJ* 2007; **335**: 1194–1199.
- 124 Richelsen B, Tonstad S, Rossner S, Toubro S, Niskanen L, Madsbad S *et al*. Effect of orlistat on weight regain and cardiovascular risk factors following a very-low-energy diet in abdominally obese patients: a 3-year randomized, placebo-controlled study. *Diabetes Care* 2007; **30**: 27–32.
- 125 Padwal R, Kezouh A, Levine M, Etminan M. Long-term persistence with orlistat and sibutramine in a population-based cohort. *Int J Obes (Lond)* 2007; **31**: 1567–1570.
- 126 Pinder RM, Brogden RN, Sawyer PR, Speight TM, Avery GS. Fenfluramine: a review of its pharmacological properties and therapeutic efficacy in obesity. *Drugs* 1975; **10**: 241–323.
- 127 Connolly HM, Crary JL, McGoan MD, Hensrud DD, Edwards BS, Edwards WD *et al*. Valvular heart disease associated with fenfluramine-phentermine. *N Engl J Med* 1997; **337**: 581–588.
- 128 Van Gaal LF, Rissanen AM, Scheen AJ, Ziegler O, Rossner S. Effects of the cannabinoid-1 receptor blocker rimonabant on weight reduction and cardiovascular risk factors in overweight patients: 1-year experience from the RIO-Europe study. *Lancet* 2005; **365**: 1389–1397.
- 129 Christensen R, Kristensen PK, Bartels EM, Bliddal H, Astrup A. Efficacy and safety of the weight-loss drug rimonabant: a meta-analysis of randomised trials. *Lancet* 2007; **370**: 1706–1713.
- 130 Torp-Pedersen C, Caterson I, Coutinho W, Finer N, Van Gaal L, Maggioni A *et al*. Cardiovascular responses to weight management and sibutramine in high-risk subjects: an analysis from the SCOUT trial. *Eur Heart J* 2007; **28**: 2915–2923.
- 131 James WPT, Caterson ID, Coutinho W, Finer N, Van Gaal LF, Maggioni AP *et al*. Effect of Sibutramine on Cardiovascular Outcomes in Overweight and Obese Subjects. *New Engl J Med* 2010; **363**: 905–917.
- 132 Kokoeva MV, Yin H, Flier JS. Neurogenesis in the hypothalamus of adult mice: potential role in energy balance. *Science* 2005; **310**: 679–683.
- 133 Lambert PD, Anderson KD, Sleeman MW, Wong V, Tan J, Hjarunguru A *et al*. Ciliary neurotrophic factor activates leptin-like pathways and reduces body fat, without cachexia or rebound weight gain, even in leptin-resistant obesity. *Proc Natl Acad Sci USA* 2001; **98**: 4652–4657.
- 134 Ettinger MP, Littlejohn TW, Schwartz SL, Weiss SR, McIlwain HH, Heymsfield SB *et al*. Recombinant variant of ciliary neurotrophic factor for weight loss in obese adults: a randomized, dose-ranging study. *JAMA* 2003; **289**: 1826–1832.
- 135 Sjostrom L. Bariatric surgery and reduction in morbidity and mortality: experiences from the SOS study. *Int J Obes (Lond)* 2008; **32**(Suppl 7): S93–S97.
- 136 Powell AG, Apovian CM, Aronne LJ. New drug targets for the treatment of obesity. *Clin Pharmacol Ther* 2011; **90**: 40–51.
- 137 Neary NM, Small CJ, Druce MR, Park AJ, Ellis SM, Semjonous NM *et al*. Peptide YY3-36 and glucagon-like peptide-17-36 inhibit food intake additively. *Endocrinology* 2005; **146**: 5120–5127.
- 138 Field BC, Wren AM, Peters V, Baynes KC, Martin NM, Patterson M *et al*. PYY3-36 and oxyntomodulin can be additive in their effect on food intake in overweight and obese humans. *Diabetes* 2010; **59**: 1635–1639.